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Detection of granulocyte antibodies: Novel technologies

Petter Höglund, MD PhD

Center for Hematology and Regenerative Medicine (HERM), Karolinska
Institutet, Department of Medicine Huddinge

Clinical Immunology and Transfusion Medicine, Karolinska University
Hospital

petter.hoglund@ki.se

petter.hoglund@sll.se



Detection of granulocyte antibodies: standard techniques

- GIFT: Granulocyte immunofluorescence test
 - LIFT: Lymphocyte immunofluorescence test
 - GAT: Granulocyte agglutination test
 - MAIGA: Monoclonal antibody-specific immobilization of granulocyte antigens

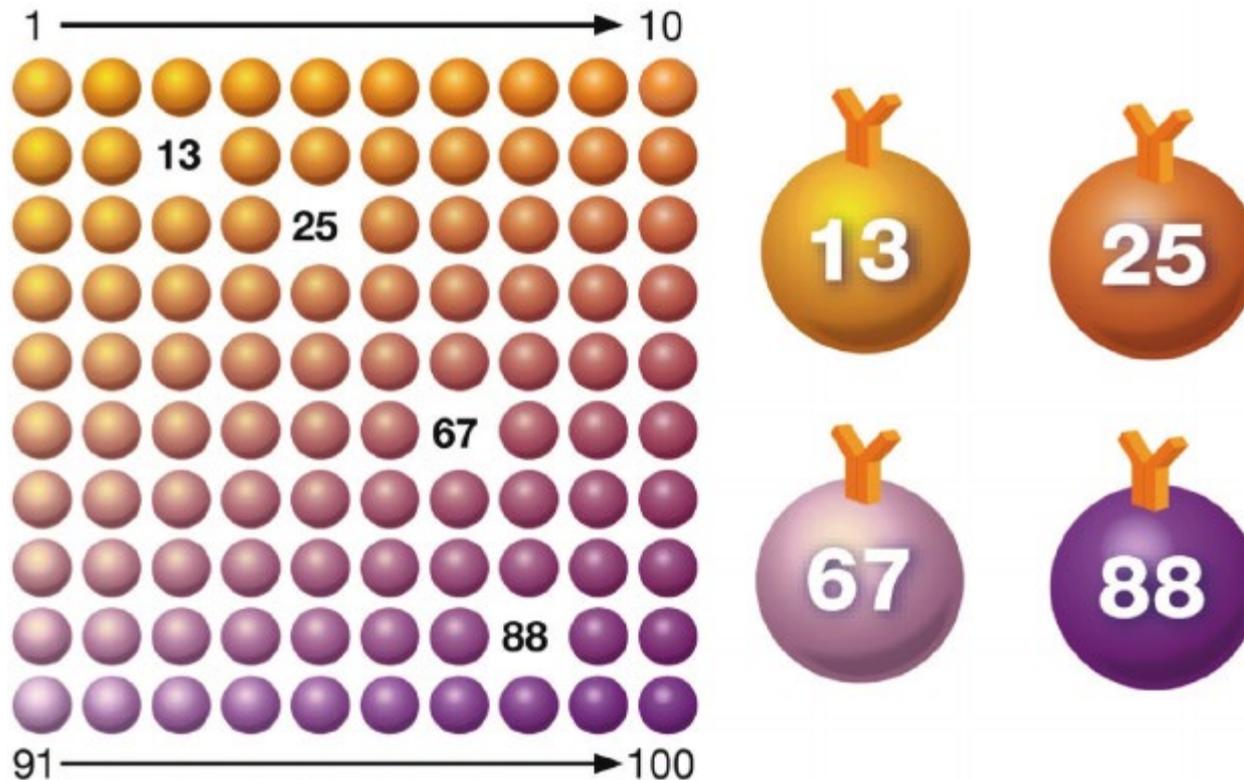
 - *Most efficient on live donor-derived granulocytes (or lymphocytes) typed for HNA-alleles*
 - *Tests with full antigen-coverage cause pressure on donors*
 - *Only MAIGA eliminates the effect of HLA-antibodies*
 - *Complicated tests requiring good health of the granulocytes and specially trained staff*
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On the wish-list

- Off-the-shelf method
 - Bulk analysis on stored samples at any time
 - Reliable antibody identification without allelic crossreactivities
 - Eliminate the need for blood donors
 - Eliminate the need for specially trained staff
 - High throughput

 - *Luminex-based technologies are promising "novel" tools*
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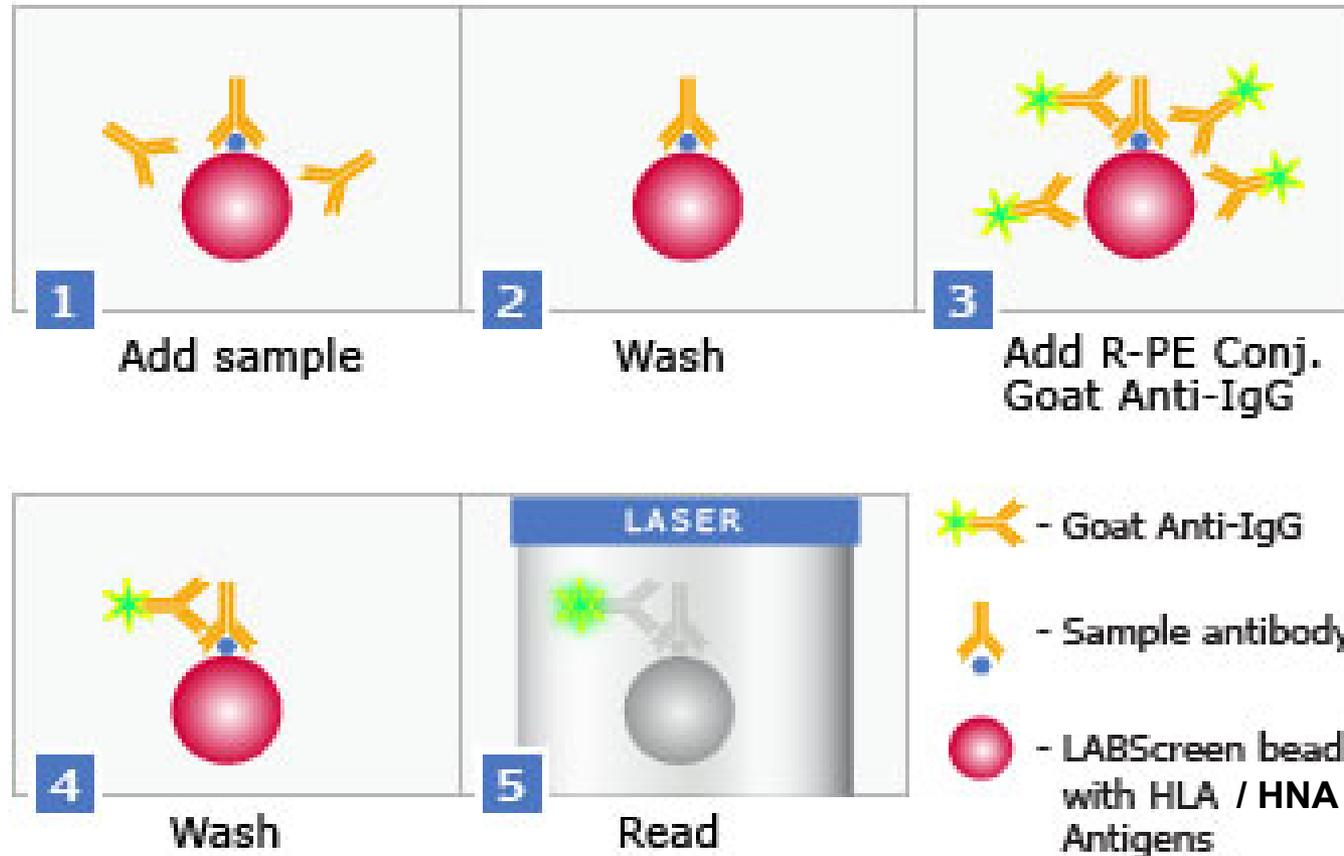
Principle: bead-technology



Each bead is individually detectable by a distinct fluorescence signal

LabScreen Multi (LSM – One Lambda)

Method



LSM setup

HLA-class I: **12 beads** with different combinations of specificities

HLA-class II: **5 beads** with different combinations of specificities

HNA: **9 beads** each expressing the following specific antigens:
HNA-1a, HNA-1b, HNA-1c, HNA-2, HNA-3a, HNA-3b, HNA-4a,
HNA-5a, HNA-5b

Performance: Incubation 1: 20 µL sample incubated with beads
Incubation 2: anti-IgG- PE

Results:
$$\text{NBG ratio} = \frac{\text{S\#N} - \text{SNC bead}}{\text{BG\#N} - \text{BGNC bead}}$$

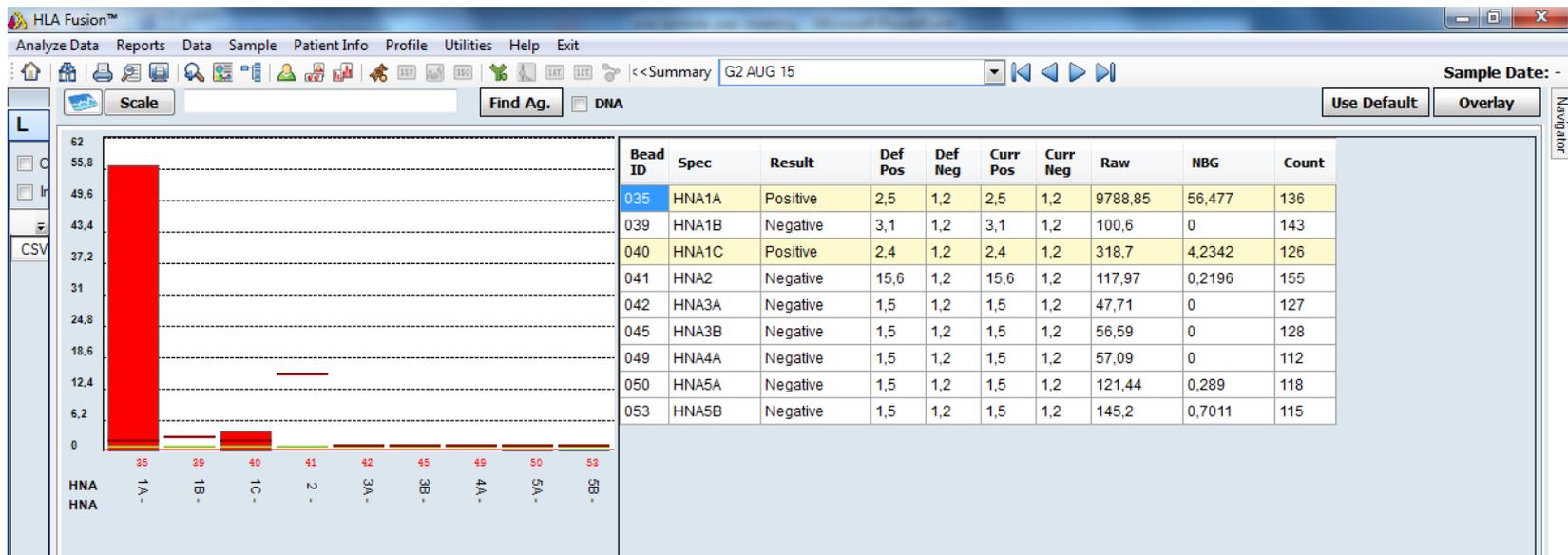
S#N
SNC bead
BG#N
BGNC bead

Sample-specific fluorescent value for bead #N
Sample-specific fluorescent value for Negative Control bead
Background NC Serum fluorescent value for bead #N
Background NC Serum fluorescent value for Negative Control bead

Alloantibodies against HNA (and HLA)

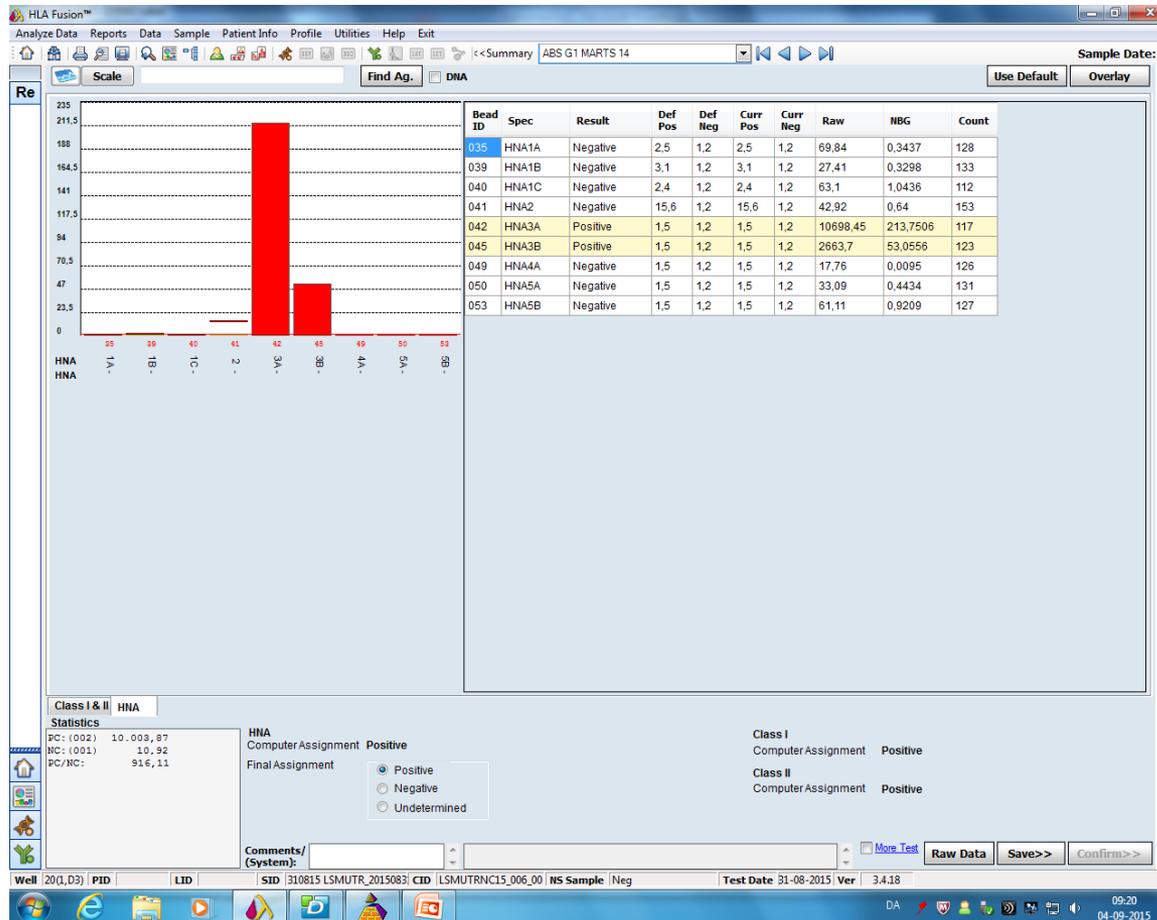
- Alloimmunization against HNA-antigens can occur during pregnancy or blood transfusion.
 - Alloantibodies against HNA-antigens (in particular anti-HNA-3 antibodies) can cause transfusion-related acute lung injury (TRALI).
 - Antibodies to foreign HLA can also cause TRALI.
 - Plasma from female donors is generally not used clinically.
 - Screening of blood donors for HLA/HNA-antibodies reduce TRALI incidence.
 - Important aspect of granulocyte antibody laboratories to identify such antibodies.
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Example: detection of anti-HNA-1a alloantibody using LSM



Courtesy of Kaspar René Nielsen, Aalborg University, Denmark

Example: detection of anti-HNA-3a alloantibody using LSM

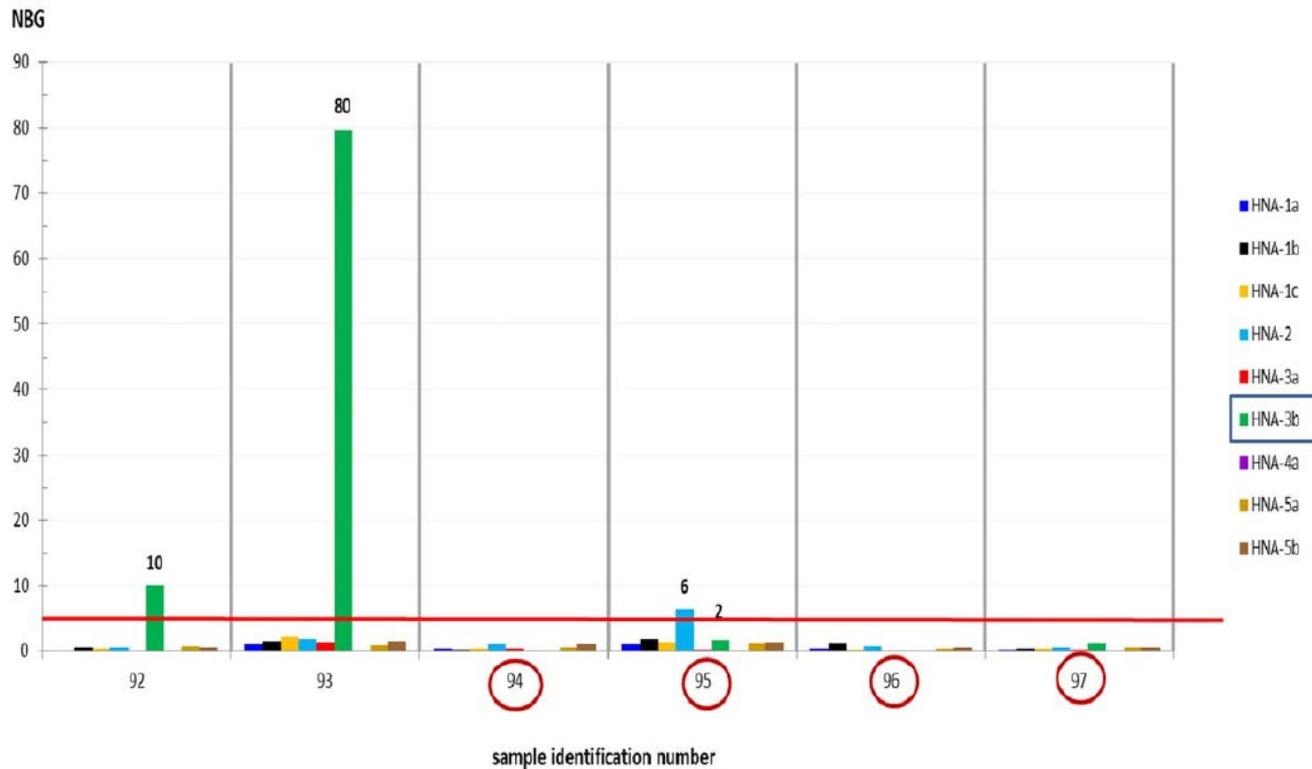


Courtesy of Kaspar René Nielsen, Aalborg University, Denmark

Performance of positive controls (alloantibodies) in LSM in our laboratory

Spec	Bead#	NIBSC HNA-1a	HNA-1b	HNA-2	HNA-3a NIBSC	HNA-3b	HNA-4a
NC	1	58	79	45	9	19	157
PC	2	6002	2226	5320	6313	4081	6145
HNA-1a	35	5928	139	140	98	57	163
HNA-1b	39	121	6665	128	71	80	168
HNA-1c	40	545	4938	124	66	49	149
HNA-2	41	117	157	23632	61	48	255
HNA-3a	42	151	177	129	11034	37	150
HNA-3b	45	176	208	158	2926	274	183
HNA-4a	49	145	66	41	17	151	9936
HNA-5a	50	127	228	143	45	131	213
HNA-5b	53	124	127	125	70	64	153

Many false negatives for HNA-3b also in published material



Schultz et al., *Transfusion*, 57, 70, 2017

LSM for alloantibody detection

- Detects almost all known HNA-1, HNA-2 and HNA-4 alloantibodies identified using other methods.
 - 10% of known HNA-3a and 67% of known HNA-3b antibodies are missed.
 - Crossreactivity is often seen between HNA-3a and HNA-3b
 - GAT is still be needed as a complement for TRALI investigations.
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LSM in patients with neutropenia: preliminary conclusions from our laboratory

- Individual cut-offs must be determined in a large number of healthy blood donors
 - LSM has a rather good correspondence with the MAIGA test for HNA-1 in a large cohort of previously analysed AIN patients.
 - Correspondence for HNA-2, HNA-4 and HNA-5 is much poorer and LSM does not detect these weaker antibodies.
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LSM – summary

- LSM is a fast, easy-to-use, high-throughput kit.
 - It tests for HLA class I, class II and HNA antibodies at the same time and uses only a very small sample volume (20 μ l).
 - Cut-offs based on samples from non-immunized male donors and (if possible) healthy children should be individualized for each lab.
 - At the Karolinska, we have performed LSM for all neutropenia investigations since February 2020. Follow-up is in progress, including stratification for adult and pediatric samples and correlation to other tests.
 - LSM has been tested also for screening assay blood donors. 2,3 % were positive for HNA antibodies and many more for HLA antibodies (close to 20%). NBG ratios were between 8 and 22. Follow-up is ongoing with repeated testing.
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Reflections on the diagnostics of autoantibodies in suspected AIN

- The presence of autoantibodies supports a diagnosis of AIN but their absence does not exclude it.
 - It is important not to mislead clinicians (and possibly delaying further investigations) by answering non-specific reaction as positive. There is no need to strive for maximum sensitivity at the cost of specificity. Better to identify too few than too many antibody-positive samples!
 - There is no "answer section" you can look up in the back of the book to find out if a specific sample contains real anti-neutrophil antibodies or not. Thus, we cannot à priori know which method most adequately reflect the clinical entity. Clinical studies are required!
 - My opinion: The best method for autoantibody detection is the one that reproducibly identifies positive results in a fraction of samples from patients with a clinical history of suspected acquired neutropenia. At the same time, the method should be cost-efficient, off the shelf, easy-to-perform and (preferrably) independent from blood donors.
 - LSM is an interesting new method, with the drawback of lower sensitivity compared to classical methods. Comparative studies are needed, including clinical follow-ups.
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